

Table S2. Strains, plasmids, and primers used in the present study.

	Designation/sequence	Reference/source/purpose
Strains		
MW2 (WT)	<i>S. aureus</i> MW2, community-associated MRSA strain	(Baba et al., 2002) ¹
$\Delta psma\beta hld$	<i>S. aureus</i> MW2 with <i>psma</i> and <i>psmβ</i> operons deleted and the <i>hld</i> start codon altered to abolish translation	(Cheung et al., 2012)
$\Delta psma\beta hld\Delta pmtR$	$\Delta psma\beta hld$ with <i>pmtR</i> gene deleted	This study
Δagr	<i>S. aureus</i> MW2 with <i>agr</i> system deleted	(Wang et al., 2007)
<i>E. coli</i> BL21 (DE3)	<i>E. coli</i> host for protein expression <i>fhuA2</i> [<i>lon</i>] <i>ompT</i> <i>gal</i> (λ DE3) [<i>dcm</i>] $\Delta hsdS$ λ DE3 = λ s <i>BamH</i> I <i>o</i> Δ <i>EcoRI</i> - <i>B</i> <i>int</i> :(<i>lacI</i> :: <i>PlacUV5</i> :: <i>T7 gene1</i>) <i>i21</i> Δ <i>nin5</i>	Life Technologies
Plasmids		
<i>pTXpmtR</i>	Constitutive expression vector expressing PmtR; tet ^R	This study
<i>pTX16</i>	Control vector for pTX _a plasmid series; tet ^R	(Wang et al., 2007)
<i>pTXpsma</i>	Xylose-inducible expression vector expressing PSMα1-4; tet ^R	(Chatterjee et al., 2013)
<i>pKX16</i>	Control vector for pKX plasmid series; kan ^R	(Chatterjee et al., 2011)
<i>pKXpsma</i>	pKX expressing PSMα1-4 upon xylose induction; kan ^R	This study
<i>pKXpsmβ</i>	pKX expressing PSMβ1-2 upon xylose induction; kan ^R	This study
<i>pKXhld</i>	pKX expressing δ-toxin upon xylose induction; kan ^R	This study
<i>pGEX-4T-1</i>	Vector for GST fusion protein cloning and expression; amp ^R	GE Healthcare
<i>pGEX-4T-1gst-pmtR</i>	Vector expressing GST-PmtR; amp ^R	This study
<i>pTX16</i>	Control vector for pTX plasmid series; tet ^R	(Peschel et al., 1996)
<i>pTXpsma1</i>	pTX expressing PSMα1 upon xylose induction; tet ^R	This study
<i>pTXpsma2</i>	pTX expressing PSMα2 upon xylose induction; tet ^R	This study
<i>pTXpsma3</i>	pTX expressing PSMα3 upon xylose induction; tet ^R	This study
<i>pTXpsma4</i>	pTX expressing PSMα4 upon xylose induction; tet ^R	This study
Primers		
pmtR-P1	ggggacaagtgtacaaaaaaggcaggcttcgttttgtcgcc atacctatagttgc	Deletion of <i>pmtR</i>
pmtR-P2	caacgtccccatcatcttctg	Deletion of <i>pmtR</i>
pmtR-P3	gtgatagggggacgttgatgaatgccatagaattaatgt	Deletion of <i>pmtR</i>
pmtR-P4	ggggaccacttgtacaagaaagctggtaggtaaaagaacttttt gagtcaagc	Deletion of <i>pmtR</i>
pmtR-for	gaggatctaggggacgttgataaaaataatttaaaaaacaatag	Cloning of <i>pmtR</i> into pTX _a
pmtR-rev	tctaacgcgttcatgtatccctcataaatgaacg	Cloning of <i>pmtR</i> into pTX _a
GST-pmtR-for	gtggaggatccgagaacctgtacttccagggttatgaaaataattttaa aaaacaatagtgtttccgatttatgaacagattaagc	Cloning of <i>pmtR</i> into pGEX-4T-1

GST-pmtR-rev	cagtcgactcatgtatccctctataaatgaacg	Cloning of <i>pmtR</i> into pGEX-4T-1
PpmtR-for	agacattcgtttaggtaccttttattg	Amplification of <i>pmtR</i> promoter
PpmtR-rev	caacgtccccctatcacttctgtatatac	Amplification of <i>pmtR</i> promoter
PpmtR-for[BTN]	[BTN]caacgtccccctatcacttctgtatatacataatatac	Amplification of biotin-labeled <i>pmtR</i> promoter
PpmtRmut1-rev	caacgtccccctatcacttctgtatatacataatatacacaatvtacac	Introduction of 1-bp mutation in <i>pmtR</i> promoter
PpmtRmut1-for	caacgtccccctatcacttccgtatatacataatatacacaatatac	Introduction of 1-bp mutation in <i>pmtR</i> promoter
M13L	gtaaaacgacggccag	Fragment analysis (Queck et al., 2008)
M13R	caggaaacagctatgac	Fragment analysis (Queck et al., 2008)
*M13L	[FAM]gtaaaacgacggccag	Fragment analysis (Queck et al., 2008)
*M13R	[FAM]caggaaacagctatgac	Fragment analysis (Queck et al., 2008)

5'RACE

Pmtrace1	aattcttaaccaaatttcaatg	RNA adapter, PCR
Pmtrace2	taccagegccatttctacc	PCR
Pmtraceseq	ctctttaaaatagaactatcttg	Sequencing

Primers and probes for qRT-PCR

pmt-for	cgttagagtcaaagtccatatgg	qRT-PCR of <i>pmtB</i>
pmt-probe	caattgcaatagtgttggtgaa	qRT-PCR of <i>pmtB</i>
pmt-rev	tggaaatgatgattgacttagaaagaa	qRT-PCR of <i>pmtB</i>

Randomized DNA fragment for EMSA control

AATCACAACTACTACCTACTGGATTAAATTATCTTATAATTCCCTGCATATAAGTTACAT
TACTTCAGCCTCTAATTGTACCCTAGCACGAAGACAAATTGTTCTACCTATATTCATCATT
GGTAAGGGATTGCATGTCCCACGTAAAACATTGTTAAACTCTTAGGTTTTGAACGATAAAAAC
TTT

¹ References

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