Microsporidia are obligate intracellular parasites that are related to fungi and characterized by a distinctive infection apparatus. Infection is mediated by resistant spores that contain a long and coiled filament that is attached to the spore apex, where the cell wall is the thinnest. When triggered, the filament rapidly bursts from the spore and everts to become a tube, following a rapid increase in internal osmotic pressure caused by aquaporin-mediated swelling (1). Microsporidia can enter host cells using two different modes, both involving the polar tube (2, 3). In the first mode, the polar tube of an external spore ejects and acts like a molecular hypodermic needle and pierces the closest host cell in its trajectory. In the second mode, the microsporidian spore is taken up by host endocytosis but then quickly evades degradation by discharging its polar tube to escape the endocytic vacuole (3). The mechanism triggering endocytosis is unclear, and the potential use of specific receptors for that purpose has not been observed (3). In both cases, the polar tube permits the transfer of the infectious cytoplasm into that of the host cell (4).

Microsporidian genomes have also attracted some attention due to their small size, but, in fact, they range in size by more than an order of magnitude (from 2.3 to >24 Mbp) (5). The smallest known microsporidian genomes, which are the smallest in any eukaryote, are found within members of the Encephalitozoonidae, a lineage that primarily infects humans and other mammals, where they can cause various mild to severe systemic diseases. Encoding roughly 2,000 genes and ranging between 2.3 and 2.9 Mbp in size (6–9), Encephalitozoonidae are extremely compact, with few introns, very short intergenic regions, and no transposable elements. Although these reduced genomes are the best studied of any microsporidian genomes, it is still unclear why they...
have been so drastically altered: reduction is sometimes linked to their obligate intracellular parasitic lifestyle (10, 11), but only vaguely, and this is not obviously consistent with the 10-fold variability in other microsporidia.

The closest known relative of members of the Encephalitozoonidae is Octosporea colligata, which infects the microcrustacean Daphnia magna (12). Infections are typically located in the anterior part of the host’s midgut, where they can reach very high intensities, with nearly every gut epithelium cell being infected (13). The consequences of infection are not as severe as those seen with some microsporidia, but the reproductive success of infected females is reduced by 20%, and they die earlier than uninfected controls (14). Infected hosts release transmission-stage parasites with their feces, and the free-floating spores are then ingested by other filter-feeding Daphnia spp. The parasite has been reported from D. magna populations in Europe and the Middle East (13, 15).

As the sister to the Encephalitozoon genome, the O. colligata genome might offer some important clues to the origin and evolution of this model for genomic reduction and compaction. Here we describe the complete sequence of the nuclear genome of O. colligata OC4 and compare it to those of its relatives from the lineage Encephalitozoonidae. We show that the structures and contents of the O. colligata Oc4 and Encephalitozoon genomes are remarkably similar, with only a few distinct chromosomal reorganizations and very limited differences in gene content, showing that the extreme reduction characterizing Encephalitozoon genomes significantly preceded the origin of the genus. The most surprising difference between the two is that the O. colligata genome has acquired a Daphnia-derived septin by horizontal gene transfer (HGT). This protein is structurally analogous to septin 7 and retains transmembrane domains, suggesting that it could be located at the spore surface. In the fungal pathogen Candida albicans, the invasion mechanism is mediated by proteins called invasins that interact with the host septin 7 to induce endocytosis of the parasite by the host endothelial microfilaments (16). The presence of a Daphnia-derived septin 7 in O. colligata could facilitate its attachment to epithelial cells of Daphnia spp. by binding directly to its N-cadherin-like surface receptors and increase the likelihood of infection either simply by maintaining proximity to the host or more directly by triggering the host’s endocytosis mechanism.

RESULTS

Genome structure of O. colligata. The assembled O. colligata OC4 genome resulted in a total of 2,290,528 bp of unique sequence distributed in 15 contigs (627× average coverage). The 12 largest O. colligata single-copy contigs are structurally similarly to those of the 11 Encephalitozoon chromosomes, although there is evidence of some large transpositions such as the equivalents of chromosomes V and IX split and attached to other loci (Fig. 1). In contrast, the remaining three small contigs (5,456 to 16,597 bp) are repeated up to four times in the genome based on their respective coverage data and correspond to subtelomeric regions found in the Encephalitozoon species that could not be linked unambiguously by PCR.

One of the main structural differences between the O. colligata and Encephalitozoon genomes pertains to the telomeric regions. In the Encephalitozoon genomes, the rRNA operons are present in the subtelomeric regions of each chromosome, for a total of at least 22 copies (6–9). In contrast, the rRNA operons are present only four times in the O. colligata genome, indicating that its subtelomeric regions are structured differently from those in Encephalitozoon species. The sharp increase in GC richness in the Encephalitozoon subtelomeric regions compared with their chromosome cores (Fig. 1) was not observed in O. colligata; however, the cores themselves display the same arcing G+C% pattern as the Encephalitozoon species (Fig. 1) (7, 9), peaking in the central portions. At 38.2%, the overall G+C content of the O. colligata genome is lower than that of Encephalitozoon species and, perhaps not coincidentally, closest to that of the invertebrate pathogen E. romaleae (Table 1).

Evolution of gene content in O. colligata and Encephalitozoon spp. To first confirm the relationship between O. colligata and Encephalitozoon spp. that has previously been inferred from small-subunit (SSU) rRNA (17), we reconstructed the phylogeny based on 104 proteins that are shared between all microsporidian species investigated. This analysis robustly confirmed the positioning of O. colligata at the base of the Encephalitozoon species (see Fig. S1 in the supplemental material) (17), a phylogenetic affiliation that is also supported by gene content and metabolic profiling (see below). O. colligata is not closely related to Hamiltosporidium tvamminensis (formerly called Octosporea bayeri), which also infects Daphnia spp. (18) and, in some parts of their species range, coinfects the same host individual as O. colligata (19). Daphnia spp. are susceptible to many microsporidian species, which are widespread across the phylogenetic tree (17, 20, 21).

Annotating all open reading frames using homology to known proteins and positional orthology with E. cuniculi resulted in a total of 1,801 discrete protein-coding genes found in the O. colligata assembly. Overall, this makes the O. colligata genome only slightly less compact than those of its Encephalitozoon relatives, with very similar coding density (0.82 genes/kb versus 0.83 genes/kb in E. cuniculi), gene content, and lack of repeats and similar intron distribution (Table 1). Most of the genes that were identified are shared with Encephalitozoon species, even in those cases where Encephalitozoon spp. differ from other microsporidia. For example, O. colligata is incapable of endogenous RNA interference and lacks the Dicer and Argonaute proteins found in the 6-Mbp + genomes of the microsporidian species Nosema ceranae, Spraguea lophii, and Trachipleistophora hominis (22–24). Unsurprisingly, the folate-related genes that were acquired by HGT in the Encephalitozoon lineage leading to E. hellem and E. romaleae (6) are also not found in O. colligata. However, the two ricin B-lectin domain-containing paralogs that are conserved across microsporidian species (22) and duplicated in tandem in the E. cuniculi strains (from ECU08_1700 to ECU08_1730) (7, 9) are also absent from the O. colligata genome. Instead, a single open reading frame (ORF) (M896_091670) that does not display any significant homology to genes encoding other proteins is located in the corresponding locus, suggesting that the ricin b-lectin paralogs were either amplified in Encephalitozoon spp. or perhaps lost or heavily modified and reduced in O. colligata.

Up to 95% of the O. colligata proteome is shared with the Encephalitozoon species (Fig. 2). In contrast, only 74% identity is found in Nosema species (a member of the sister group corresponding to the O. colligata/Encephalitozoon clade), with percentages decreasing rapidly as one moves further away in the tree, for an averaged pairwise proportion of 50% (highest, 100%; lowest,
16%). However, these low percentages do not necessarily mean that the genes are nonhomologous, since they may simply reflect the high rate of sequence divergence occurring in microsporidia. None of the *O. colligata* proteins that are absent from *Encephalitozoon* spp. have been found in other microsporidia, and, with the exception noted below, none have identifiable functions. These unique protein-coding genes are not restricted to subtelomeric regions; many are inserted within the cores of the *O. colligata* chromosome between genes arrayed in otherwise syntenic fashion with other *Encephalitozoon* spp. However, in 14 cases that included the M896_091670 gene inserted in lieu of the b-lectin genes, positional orthologs that are, however, dissimilar in sequences are found between the *O. colligata* and *Encephalitozoon* cores, suggesting that the corresponding genes may not be unique to *O. colligata* but may rather be divergent beyond recognition.

About 58% of the *O. colligata* proteome can be assigned putative functions, consistent with other microsporidia, for which 29% to 61% of the proteome has identifiable functions. The relative distributions of functions across metabolic pathways for identifiable proteins in microsporidia are similar across most species (Fig. 2; see also Table S1 in the supplemental material), with an elevated proportion of proteins involved in amino acid and carbohydrate metabolism in the genomes from basal *Rozella* and *Mitosporidium* species, which is congruent with their closer position-
ing to other nonpathogenic fungi. Surprisingly, Nosema bombycis also displays an elevated proportion of proteins involved in carbohydrate metabolism, perhaps resulting from its recent abundant genomic duplications (25). The human pathogen Enterocytozoon bieneusi displays a large proportion of genes for amino acid biosynthesis, signal transduction, and translation-related components, the latter of which represent the highest proportion in any sequenced microsporidian, 45% more than the second highest tally in the gene-rich genomes of Mitosporidium and Cryptomyzocota Rozella species (Fig. 1; see also Table S1).

To investigate the levels of divergence between the *O. colligata* and *Encephalitozoon* protein-coding genes, we aligned all shared orthologs and calculated the nucleotide diversity within *Encephalitozoon* species (Pi) and between *O. colligata* and *Encephalitozoon* spp. (K1[JC]) (see Table S2 in the supplemental material). In both absolute [K1[JC]] and relative [K1[JC]/Pi] rates, polar tube protein 2 (PTP2; M896_080490) stands out as one of the fastest-absolute [K1(JC)] rates and relative [K1(JC)/Pi] rates, polar tube spp. [K1(JC)] (see Table S2 in the supplemental material). In both peptide has been preserved, but the 8-amino-acid-long lysine-rich identity at most over the aligned regions. Its N-terminal signal alitozoon orthologs at the amino acid level, with 25% pairwise identity at most over the aligned regions. Its N-terminal signal peptide has been preserved, but the 8-amino-acid-long lysine-rich identity at most over the aligned regions. Its N-terminal signal peptide has been preserved, but the 8-amino-acid-long lysine-rich domain located inside the core of the *Encephalitozoon* PTP2 proteins (26) is only barely recognizable in *O. colligata* (KPKKKKSK versus VPVKEKAR, respectively). Other highly variable genes between the two genera with identifiable functions include an endochitinase (M896_021680; orthologous to ECU09_1320) involved in spore wall maintenance, a phosphoacetylglucosamine mutase (M896_010540; orthologous to ECU01_0650) involved in carbohydrate metabolism, and a ubiquitin carboxyl-terminase hydrodase (M896_060900; orthologous to ECU06_0910) involved in ubiquitin conjugation. *O. colligata* contains two additional copies of another ubiquitin carboxyl-terminase hydrodase orthologous to ECU03_0580. One of these, M896_031130, is syntenic with its *Encephalitozoon* orthologs, but the other, M896_051260, is paralogous and likely arose from intragenomic duplication. The M896_031130/ECU03_0580 orthologs are not particularly divergent, but the paralogous M896_051260 is highly derived.

**Horizontal gene transfer of host-derived septin gene to *O. colligata***. In addition to subtelomeres, the *O. colligata* genome has three other regions of unusually low G+C content, but these are not common to *Encephalitozoon* spp. or other microsporidia (Fig. 1 and 3). In all three cases, the overall G+C percentage is about 10% lower than the content in the surrounding regions. Two of the three segments (on contigs 3 and 8) are inserted within blocks of genes that are otherwise syntenic with the *Encephalitozoon* genomes, whereas the third (on contig 2) is located at one of the junctions of a major intrachromosomal transposition involving chromosome 2 (Fig. 3). We confirmed the assembly of all three regions by PCR and Sanger sequencing, excluding potential assembly errors.

The presumed transposition breakpoint is relatively gene poor, but the other two regions are of comparable density to that of the genome as a whole and encode several potential open reading frames (Fig. 3). All but a few these ORFs are of unknown function. M896_020810 has low similarity to magnesium transporters, and M896_051300 and M896_051310 feature indistinct coiled coil frames (Fig. 3). All but a few these ORFs are of unknown function. M896_020810 has low similarity to magnesium transporters, and M896_051300 and M896_051310 feature indistinct coiled coil motifs. In contrast, however, the *O. colligata* M896_080490 protein located within the low-GC region on chromosome 8 is homologous to genes encoded in the genomes of *Daphnia pulex* (E value of 4e-53 with DAPPUDRAFT_2014173; 60% similarity over 256 aligned amino acid residues) and *Daphnia magna* (E value of 2e-50; 63% similarity over 287 aligned amino acid residues) (reference 27 and unpublished data). Phylogenetic inferences derived from the alignment of M896_080490 with similar sequences from the NCBI NR database (E value BLASTP cutoff value of 1e-40) cluster this protein firmly within clades of *Daphnia pulex* paralogs (Fig. 4). The *D. pulex* and *D. magna* genomes contain large AT-rich stretches, suggesting that the AT-rich seg-

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**TABLE 1** General features of the *O. colligata* and *Encephalitozoon* genomes

<table>
<thead>
<tr>
<th>Feature</th>
<th><em>O. colligata</em> OCA</th>
<th><em>E. cuniculi</em> GB-M1</th>
<th><em>E. intestinalis</em> ATCC 50506</th>
<th><em>E. hellem</em> ATCC 50504</th>
<th><em>E. romaleae</em> SJ-2008</th>
</tr>
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<tbody>
<tr>
<td>No. of chromosomes</td>
<td>10</td>
<td>11</td>
<td>11</td>
<td>11</td>
<td>11</td>
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<tr>
<td>Estimated total genome size (Mb, )</td>
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<td>2.3</td>
<td>2.5</td>
<td>2.5</td>
</tr>
<tr>
<td>Assembled-genome size (Mb, )</td>
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<td>2.2</td>
<td>2.3</td>
<td>2.2</td>
</tr>
<tr>
<td>Genome coverage (%)</td>
<td>77</td>
<td>86</td>
<td>96</td>
<td>92</td>
<td>88</td>
</tr>
<tr>
<td>G+C content (%)</td>
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<td>47</td>
<td>41.4</td>
<td>43.4</td>
<td>40.3</td>
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<tr>
<td>Gene density (gene/kbp)</td>
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<td>0.63</td>
<td>0.91</td>
<td>0.89</td>
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<td>Mean gene length (bp)</td>
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<td>1041</td>
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<td>1018</td>
<td>1061</td>
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<tr>
<td>Mean intergenic length (bp)</td>
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<td>166</td>
<td>100</td>
<td>106</td>
<td>130</td>
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<tr>
<td>No. of SSU-LSU RNA genes</td>
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<tr>
<td>No. of mcRNAs</td>
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<td>14</td>
<td>11</td>
<td>NA</td>
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<tr>
<td>No. of tRNAs</td>
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<td>46</td>
<td>46</td>
<td>46</td>
<td>46</td>
</tr>
<tr>
<td>No. of tRNA introns (sizes in bp)</td>
<td>2 (11, 42)</td>
<td>2 (12, 41)</td>
<td>2 (12, 41)</td>
<td>2 (12, 41)</td>
<td>2 (12, 41)</td>
</tr>
<tr>
<td>No. of splic. introns (size range in bp)</td>
<td>30 (23–77)</td>
<td>36 (23–76)</td>
<td>36 (23–76)</td>
<td>36 (23–76)</td>
<td>36 (23–76)</td>
</tr>
<tr>
<td>Predicted no. of ORFs</td>
<td>1820</td>
<td>2010</td>
<td>1944</td>
<td>1928</td>
<td>1835</td>
</tr>
<tr>
<td>Predicted no. of ORFs</td>
<td>1820</td>
<td>2010</td>
<td>1944</td>
<td>1928</td>
<td>1835</td>
</tr>
</tbody>
</table>

a The *E. cuniculi* and *E. romaleae* values from Pombert et al. (6). The *E. intestinalis* and *E. hellem* values are from the 2014 accession updates.

b The numbers of chromosomes for *O. colligata* were estimated based on the chromosomal reorganizations observed with the *Encephalitozoon* species.

c The numbers of small subunit and large subunit (SSU-LSU) rRNA genes were inferred based on their overall coverage relative to other genes.

d NA, the mcRNAs in *E. cuniculi* and *E. romaleae* were not assessed. They are likely similar in number and content to those found in *O. colligata*, *E. intestinalis*, and *E. hellem*. 

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FIG 2. Pairwise distribution and metabolic profiling of the microsporidian proteome. The phylogenetic positions in the cladogram are derived from our phylogenetic inferences (see Fig. S1 in the supplemental material). Nodes recovered in all of the bootstrap replicates are indicated by asterisks, whereas values indicate levels of support for the corresponding nodes. Assembled-genome sizes are indicated in Mb between each branch and the corresponding microsporidian taxa. In the adjacent heat map, light and dark colors indicate the percentages of proteins from the branching species that are shared with the species indicated on top. Darker colors indicate more shared proteins.

The Ordospora colligata Genome; of Reduction and HGT January/February 2015 Volume 6 Issue 1 e02400-14
tifs, is exposed outside the membrane (see Data S1 at https://github.com/FP-Laboratory/mBio-2015/). Two domains, spanning residues 1 to 221 and residues 222 to 794, were predicted using the Ginzu hierarchical screening method. The first domain has weak (Ginzu confidence of 0.0735) and likely spurious structural homology with DNA cleavage/binding domains (2NRRA_101) and the hemolysin binding component of Bacillus cereus toxins (2NRJA), but the second has a much stronger match (Ginzu confidence of 0.607009) as a septin hydrolase/GTPase, similar to human septin 7 (Protein Data Bank [PDB] entry 2QAG.1.C). No signal peptide suggestive of secretion was predicted. Interestingly, two other septins (M896_011310 and M896_100270) were found to be among the fastest-evolving genes between the O. colligata and Encephalitozoon species, with K(JC)/Pi ratios of 2.70 and 2.66, respectively.

**DISCUSSION**

The Encephalitozoon genus has been held as a model for extreme genome reduction and compaction, and the patterns left by these processes over the evolution of the genus have been well studied (6, 8, 9, 28). Explanations for why these genomes are so compact are less clear, but the 10-fold variation in genome size in microsporidia suggests that it is not intrinsically related to intracellular parasitism. Here, we show that the extreme reduction in the Encephalitozoon genome actually predates the origin of the genus and is common to the Daphnia-infecting sister lineage represented by O. colligata. Indeed, in terms of gene density, content, and overall structure, the O. colligata genome is hardly distinguishable from those of Encephalitozoon spp. The impressive level of synteny observed between the two genera strongly argues against independent rounds of reductions, such that the genome of the last common ancestor of the O. colligata and Encephalitozoon species was almost assuredly similar in every aspect.

Of the differences in gene content we do describe, however, the septin 7 homologue stands out as being of particular functional and evolutionary interest. Obligate intracellular parasites have limited opportunities to exchange genetic material, given that their sheltered environment offers few opportunities to exchange genes with organisms other than the host. Turning a host’s proteome against itself is a strategy that has been used by both prokaryotic and eukaryotic pathogens throughout the course of evolution, although clear-cut cases of host-to-parasite HGT in eukaryotes are still rare in general (29, 30). Horizontal gene transfer has been shown in microsporidians, despite their genome-reductionist tendencies, and some of their newly acquired genes have obvious possible benefits with respect to infection or survival outside their host (31, 32). Only a single case of HGT from an animal host has been reported in microsporidia, however (6, 33), and here we show that a septin 7 in O. colligata not only is animal

**Figure Legend Continued**

higher levels of conservation; conservation is reduced as we move further across the phylogenetic tree. For each species, the total number of predicted genes indicated in the center of the figure. "uORF" (unknown ORF) refers to the number of proteins for which no gene ontology (GO) could be ascribed. pfUNC refers to predicted proteins with at least one associated GO in InterProScan 5 analyses. On the right, the metabolic profiles (in percentages) derived from the detected GO are categorized according to KEGG pathways and drawn to scale. Ami, amino acid metabolism; Car, carbohydrate metabolism; Cgd, cell growth and death; Cmc, cell motility cytoskeleton; Ene, energy metabolism; Fsd, folding, sorting, and degradation; Lip, lipid metabolism; Mem, membrane transport; Mcv, metabolism of cofactors and vitamins; Mtp, metabolism of terpenoids and polyketides; Mis, miscellaneous; Nuc, nucleotide metabolism; Rep, replication and repair; Sig, signal transduction; Tsc, transcription; Tsl, translation; Tsc, transport and catabolism. N. antheraeae, Nosema antheraeae; N. apis, Nosema apis; V. cornaeae, Vittaforma cornaeae; A. locustae, Antonospora locustae; A. algerae, Annaliicia algerae; E. aedis, Edhazardia aedis; V. culicis floridensis, Venera culicis floridensis; N. parisii, Nematocida parasites; N. sp. 1, Nematocida sp. 1; M. daphniae, Mitosporidium daphniae; R. allomyza, Rozella allomyza.
FIG 4  Maximum likelihood phylogenetic inferences of *O. colligata*’s septin 7 (M896_080490) and similar protein sequences. All protein sequences displaying similarity greater than the selected cutoff (*E* value cutoff 1E to 40; accession date, 20 August 2014) were retrieved from the NCBI NR database. The best ML tree (LG4) is shown here. The major nodes retrieved in all bootstrap replicates are indicated by asterisks, whereas numbers indicate the corresponding levels of bootstrap support (values lower than 60 are not shown). *Daphnia* paralogous clades are color coded with a bluish gradient and labeled A to D.

derived but is apparently closely related to homologues in *D. pulex* and *D. magna* genomes. The directionality of this transfer is clear; no homologues are found in other fungi, and the transfer is likely recent as well. The horizontal transfer seems to have occurred after the split between the *O. colligata* and *Encephalitozoon* spp. and to have resulted in the acquisition of additional transmembrane domains since its origin. The alternative, acquisition by their common ancestor and then loss in the members of the *Encephalitozoonidae*, seems unlikely given their host range histories.

The function of the *O. colligata* septin is unknown, but other fungal pathogens offer intriguing possibilities related to cell division and compartmentalization (34). Intuitively, cell division proteins form valuable acquisition targets for vertically transmitted parasites to modulate the reproductive cycle of hosts. Here, however, the function of the HGT-acquired septin is unlikely to be involved in cell division: unlike the microsporidian *Daphnia* pathogen *Pereziella diaphanosoma*, which can infect both gut and reproductive organs (12), *O. colligata* is a strictly horizontally transmitted gut epithelium pathogen that has never been encountered in reproductive organs. Thus, a secreted septin would not disrupt host reproduction. There is also no evidence that this HGT-acquired septin is secreted and, considering the evolutionary distance between the parasite and its hosts, it is unlikely that this protein would play a role in *O. colligata*’s own cell division. One could, however, imagine a function in compartmentalization, for example, inducing endocytosis by the host. *Encephalitozoon* species can invade their host by endocytosis (2, 3), and in the fungal pathogen *Candida albicans*, the endocytic invasion is initiated by proteins called invasins that interact with the host septin 7, a major effector (35), which initiates a molecular cascade, ultimately inducing the uptake of the pathogen (16). The septin acquired by *O. colligata* is structurally analogous to septin 7 and features additional transmembrane motifs, suggesting that it could be localized at the proteinaceous exosporium. An externally exposed septin could camouflage *O. colligata* and bypass the need for invasins by binding to cell surface N-cadherins of *Daphnia* spp., by recruiting other components of the host septin 2/6/7 complex, or by interacting directly with the microfilaments of the gut epithelium to facilitate entry into the host by endocytosis. Alternatively, a surface septin could also facilitate infection by simply helping to keep the parasite in close proximity to the host cell surface.

Unfortunately, little is known about the infection process in *O. colligata* (or in many other microsporidia for that matter), so more direct evidence is required to determine if *O. colligata* is capable of infection by self-induced endocytosis and, if so, if the HGT-acquired septin is surface localized and facilitates this process. There is no guarantee that the gene has remained functional since being acquired by HGT, and while the septin core appears to have been conserved in *O. colligata* (see Fig. S2 in the supplemental material), the long branch it displays in phylogenetic analyses (Fig. 4) suggests that it is evolving at a fast pace. In any case,
because *Encephalitozoon* species are capable of infecting their host by endocytosis (3) and yet lack the septin 7 found in *O. colligata*, we infer that this gene is not essential to the underlying mechanism. However, considering the number of diverse microsporidian species that infect *Daphnia* spp., the presence of a functional septin 7 in *O. colligata* could confer to it a competitive advantage over its parasitic relatives. The distribution of this gene in related taxa is also of interest for efforts to help determine how recent the HGT was: it may have originated relatively early and been lost in *Encephalitozoon* spp. or may be present within only a limited number of genotypes.

**MATERIALS AND METHODS**

**Tissue culture and DNA purification.** *O. colligata* isolate OC4 was cultured in *Daphnia magna* clone ELK1-1 (England-LadyKirk-pond 1) (36) and used for further laboratory procedures. Approximately 1,000 female hosts infected with *O. colligata* OC4 were homogenized in 10 mM Tris-HCl (pH 7) and then filtered sequentially through a 40-μm-pore-size nylon mesh and an 8-μm-pore-size cellulose nitrate membrane adapted to a syringe. The filtrate was centrifuged at 4,000 rpm for 10 min and resuspended in 10 mM Tris-HCl (pH 7). The solution, containing spores and other tissue debris, was centrifuged in 60% Percoll (Sigma Aldrich) at 14,000 rpm for 5 min, and the pellet was washed 3 times with 10 mM Tris-HCl (pH 7). The solution, containing spores and other tissue debris, was centrifuged in 60% Percoll (Sigma Aldrich) at 14,000 rpm for 5 min, and the pellet was washed 3 times with 10 mM Tris-HCl (pH 7) to obtain a clear spore solution. Spores were incubated with lysozyme (Sigma Aldrich) (2.5 mg/ml) at 37°C for 1 h to lyse contaminant bacteria. An additional step of contaminant cell lysis was performed by adding a lysis buffer (1% SDS; 2% Triton X-100; 1 mg/ml protease K; 10 mM Tris-HCl; 1 mM EDTA; 100 mM NaCl; pH 7.0) to the mixture and incubating it at 56°C for 1 h. The lysate was centrifuged at 14,000 rpm to recover the spores, which were treated with DNase I (Sigma Aldrich) at 37°C overnight to eliminate contaminating DNA. The enzyme was inactivated with EGTA (50 mM) at 95°C for 30 min, and the spores recovered by centrifugation were frozen and thawed several times.
**Phylogenetic analyses.** For phylogenomic inferences, the microsporidian protein sequences were retrieved from the MicrosporidiaDB (62), SilkPathDB (http://silkpathdb.swu.edu.cn/silkpathdb/), and GenBank databases. Orthologous sequences were identified by BLASTP searches at an E value cutoff of 1E-20 using the *O. colligata* proteins as queries. Orthologs were aligned with MAFFT L-INS-I (45), and the ambiguous positions in the resulting alignments were filtered out with BMGE (63) using the default parameters. Maximum likelihood (ML) inference analyses were performed with PHYLML 3.0 (64) using the LG model of amino acid substitutions with four gamma categories. A total of 100 bootstrap replicate experiments were performed. Bootstrap replicates were generated with Seqboot and node percentages calculated with Consense from the PHYLIP 3.695 package (65). For horizontal gene transfer inference determinations, the M896_080490 orthologous and paralogous sequences were retrieved from the NCBI nonredundant (NR) database (accession date, 20 August 2014) using BLASTP searches with an E value cutoff of 1E-40. The retrieved sequences were aligned with MUSCLE (66), and the resulting alignment was filtered with BMGE using the default parameters. ML inference and bootstrap replicate experiments were performed as described above.

**Pairwise distribution and metabolic profiling.** For each species, local protein databases were generated using MAKEBLASTDB from the NCBI BLAST+ 2/2/28 package. The presence or absence of genes in comparisons between species was determined by evaluating pairwise BLASTP hits (E value cutoff, 1e-10) for all possible combinations. Metabolic profiles were inferred from InterProScan 5 (50) analyses performed on each protein data set; for each protein, the gene ontologies retrieved were filtered to remove duplicates and concatenated into higher hierarchies derived from the KEGG orthology pathways using custom Perl scripts.

**Nucleotide diversity.** The *O. colligata* protein-coding genes and their *Encephalitozoon* orthologs were aligned by codon comparisons performed with MACSE (67). For each alignment, the nucleotide diversity (Pi) between *Encephalitozoon* species and their divergence from *O. colligata* [K1(C)] were inferred using the polymorphism and divergence in functional regions tool implemented in DnaSP 5/10/01 (68) with *O. colligata* as the outgroup.

**Accession numbers.** The *O. colligata* data were released in the NCBI database under BioProject PRJNA210314, BioSample SAMN02867507, and accession number JOKQ00000000.

### SUPPLEMENTAL MATERIAL

**Supplemental material for this article may be found at http://mbio.asm.org/lookup/suppl/doi:10.1128/mBio.02400-14/-/DCSupplemental.**

- Figure S1, PDF file, 0.1 MB.
- Figure S2, PNG file, 0.2 MB.
- Table S1, XLSX file, 0.01 MB.
- Table S2, XLSX file, 0.5 MB.

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